Cat. No: P2101c, 100 rxns/20μl-rxn
P2102c, 500 rxns/20μl-rxn

Power Green qPCR Mix with ROX

For research use only

Components

Component	P2101c	P2102c
2X Power Green qPCR Mix	1 ml	1 ml × 5
50X ROX Reference Dye H*	40 μl	200 μl
50X ROX Reference Dye L*	40 μl	200 μl
Nuclease-free Water	1 ml	1 ml × 5

* ROX Reference Dye H and ROX Reference Dye L are suitable for fluorescence quantitative PCR instruments requiring high and low concentrations of ROX calibration respectively. It is recommended that 40 μL 50X ROX Reference Dye H or L be premixed into 1ml 2X SYBR Green qPCR Mix. The final working concentration of ROX is 1X.

Storage

This mix can be stored for 2 months at 4°C and protected from light. For longer storage, it should be kept at -20°C and protected from light. The ROX Reference Dye should be kept at -20°C and protected from light.

Description

Power Green qPCR Mix is a ready-to-use, 2X concentrated mix that contains all the reagents (except template and primers) needed for the real-time qPCR in the SYBR Green I detection format. This product is compatible with most manufacturers' real-time fluorescent quantitative PCR instruments such as Applied Biosystems, Roche, Bio-Rad, Eppendorf, Corbett and so on.

The Hotstart Taq DNA Polymerase in the mix is modified by antibody. When the temperature below 45°C, the activity of the modified polymerase will be strongly suppressed, only when the temperature reached 72°C, it can be activated. This technology helps to reduce non-specific amplification and primer dimer effectively.

The reaction system of this product can be prepared at room temperature without an ice box. The prepared PCR reaction system can be placed at room temperature for 24 hours and the amplification efficiency remains unchanged.

Applications

- Gene expression analysis
- Low-copy gene detection
- Microarray validation
- Gene knockdown validation

Features

- Compatible with many Real-time systems whether requires ROX reference dye to calibration or not
- Exceptional specificity with hot-start mechanism
- Tight reproducibility in Ct values over a broad dynamic range
- Universal instrument compatibility

Table of Instrument Guide

Instruments	Final Conc. of ROX
ABI PRISM 7000/ PRISM 7700/ 7300/ 7900HT/ StepOne/ StepOnePlus/ GeneAmp 5700	500 nM，high ROX
ABI 7500/ 7500 Fast/ ViiA 7/ QuantStudio 6/7/12K Flex; Agilent Stratagene Mx3000P/ Mx3005P/ Mx4000	50 nM，low ROX
Bio-Rad CFX96/ CFX384/ iQ/ iQ5; MJ Research Opticon 2/ Chromo 4; Roche LightCycler 480/ 96; Corbett Rotor Gene G/ Q/ 3000/ 6000; Thermo PikoReal 96; Eppendorf MasterCycler ep realplex; Cepheid Smart Cycler	No ROX

Protocol

Note: Please follow the procedures outlined in the manual of each respective instrument.

1. Preparation of reaction solution (Take the ABI StepOnePlus as an example)

Add the following reagents to the proper thermal cycler reaction tube or plate on ice:

Component	20-μl rxn	Final Conc.
2X Power Green qPCR Mix	10 μl	1X
Forward Primer (10μM)	0.4 μl	0.2μM
Reverse Primer (10μM)	0.4 μl	0.2μM
50X ROX Reference Dye H	0.4 μl	1X
Template DNA	variable	0.05-5ng/μl
Water, nuclease-free	to 20 μl	–

Note:

- The primer concentration can be further optimized. The optimal range for primers is 0.1~1μM.
- Prepare according to the recommended volume of each instrument.
- Use 1-10ng cDNA or 10-100ng gDNA for each reaction.
- Users can increase the amount of the the qPCR Mix when using low-copy gene as template.
- Users can reduce the amount of the qPCR Mix, if the melting curve comes with impure peaks.

2. Setup the plate

Transfer the reaction mixture to PCR tubes/plates. Reaction volumes can be reduced to 10 μl if the instrument supports a low volume system.

Cap or seal the reaction tubes/plates then centrifuge briefly to spin down the contents and eliminate any air bubbles.

3. Preform qPCR using the following thermal cycling condition

Set the thermal cycling conditions using default PCR thermal cysling conditions specified in the following tables according to the instrument cycling parameters and melting temperatures of the specific primers.

2-step PCR mode:

Initial Denaturation	95ºC	1 min	Holding Stage
Denaturation	95ºC	5 sec	Cycling Stage 40 Cycles
Annealing & Extension	60ºC [1]	20~60 sec [2]	
Melting curve analysis (optional) [3]			

3-step PCR mode:

Initial Denaturation	95ºC	1 min	Holding Stage
Denaturation	95ºC	5 sec	Cycling Stage 40 Cycles
Annealing	55ºC [1]	30 sec	
Extension	72ºC	30~60 sec [2]	
Melting curve analysis (optional) [3]			

Note:

[1] To improve the specificity, the annealing temperature can be increased. Set up signal collection in the Annealing&Extension or Extension phases respectively.

[2] Consider the instrument type when setting the extension time. If the amplification efficiency needs to be improved, the extension time can be increased.

[3] Different instruments have different melting curve acquisition procedures, generally according to the default melting curve acquisition procedures of the instrument.

4. Analyze the results

Data analysis varies depending on the instrument used. Please refer to your instrument user guide for information.

Important Notes

Template

Genomic DNA, plasmid DNA, or cDNA can be used as template. For optimal quantitative results use up to 20ng of genomic DNA or plasmid DNA per 20 µl reaction (for smaller volumes, the amount of template should be decreased equivalently). Using greater amounts of template may reduce the maximum fluorescence signal and linearity of standard curves due to binding of the SYBR® Green I dye to the template. For two-step RT-PCR, use either undiluted or diluted cDNA generated from up to 1µg of total RNA. The volume of the cDNA added (from the RT reaction) should not exceed 10% of the final PCR volume (e.g., for a 20 µl qPCR reaction, use up to 2.0 µl of undiluted cDNA).

Primers

Careful primer design and purification (HPLC-purified primers are recommended) is particularly important in order to minimize loss in sensitivity due to the production of nonspecific amplification products in SYBR® Green I-based qPCR. This effect becomes more prominent at low target concentrations. To maximize the sensitivity of the assay, use the lowest concentration of primers that can be used without compromising the efficiency of the PCR reaction (50-400nM of each primer). For optimal results, design primers that amplify PCR products 60-400 bp in length. The primers should exhibit a melting temperature (Tm) of approximately 60ºC. We recommend designing primers specifically for amplification of cDNA derived from mRNA. This prevents amplification of contaminating genomic DNA and inaccurate quantification of mRNA.

Melting Curve Analysis

Following real-time qPCR, melting curve analysis should always be performed to identify the presence of primer-dimers and analyze the specificity of the reaction. Program your thermocycler according to the instructions provided.

Quality Control

The absence of endodeoxyribonucleases, exodeoxyribonucl- eases and ribonucleases is confirmed by appropriate quality tests. Functionally tested in amplification of a single-copy gene from human genomic DNA.

Product Use Limitations

Power Green qPCR Mix with ROX is sold exclusively for research purposes and *in vitro* use. Neither the product, nor any individual components, was tested for use in diagnostic applications or for drug development, nor is it suitable for administration to humans or animals. Please refer to the MSDS, available upon request.