

FS™ Kit

#P3072, 5 ml

Contents:

2× FS™ Reaction Mix	1 ml × 5
FS™ Taq DNA Polymerase (2.5U/μl)	160 μl

Store at -20°C

For research use only.

In total 6 vials.

Description

FS™ Kit contains FS™ Taq DNA Polymerase and a premixed, ready-to-use FS™ Reaction Mix, which contains dNTPs, Mg²⁺ and other components at optimal concentrations for efficient amplification of DNA templates by PCR. To prepare the final PCR, only need to add primers and template DNA. The reaction Mix has higher amplification sensitivity due to optimized components and enhancer.

FS™ Taq DNA Polymerase is the latest generation Taq-based DNA polymerase developed by GDSBio. It possesses high amplification efficiency as Taq DNA Polymerase does, and fast elongation ability as KOD polymerase does, can be used in various kinds of PCR. The FS™ PCR Buffer is designed for FS™ Taq DNA Polymerase, can be used in fast amplification reaction. The elongation rate of FS™ Taq DNA Polymerase can be as fast as 3kb/min. It can shorten the amplification time by 2/3. It has 5' to 3' polymerase activity, but lacks of 3' to 5' exonuclease activity, that results in a 3'-dA overhangs PCR product.

Unit Definition

One unit is defined as the amount of the enzyme required to catalyze the incorporation of 10 nmole of dNTPs into an acid-insoluble form in 30 minutes at 70°C using hering sperm DNA as substrate.

PRODUCT USE LIMITATION.

This product is developed, designed and sold exclusively for research purposes and in vitro use only. The product was not tested for use in diagnostics or for drug development, nor is it suitable for administration to humans or animals.

Storage Buffer

20mM Tris-HCl (pH8.0), 1mM DTT, 0.1mM EDTA, 3.2mM MgCl₂, 100mM KCl, 0.1% Triton X-100, 0.1% TW 20, 0.2mg/ml BSA, 50% Glycerol

Composition of the 2× FS™ Reaction Mix

2× FS™ Buffer, 0.4mM dNTPs, 3.2mM MgSO₄, 0.02% bromophenol blue.

FS™ Reaction Mix is a proprietary formulation optimized for robust performance in PCR.

Applications

- High throughput PCR
- Long and complex template PCR
- Routine PCR with high reproducibility
- Generation of PCR products for TA cloning

Features

- **Convenient:** only primers and template DNA are added when prepare final PCR
- **High yields** of PCR products with minimal optimization
- **High efficiency:** saving your time by simplifying the process
- **Reproducible:** lower contamination and pipetting error risk
- **Higher sensitivity and fast** compared to conventional Taq DNA Polymerase
- **Flexible:** the amount of polymerase is flexible and controllable.

Basic PCR Protocol

All solutions should be thawed on ice, gently vortex and briefly centrifuge.

1. Add the following components to a sterile microcentrifuge tube sitting on ice:

Reagent	Quantity	Final concentration
2× FS™ Reaction Mix	25 µl	1×
Forward Primer	variable	0.4-1µM
Reverse Primer	variable	0.4-1µM
Template DNA	variable	10pg-1µg
FS™ Taq DNA Polymerase (2.5U/µl)	0.5-1 µl	1.25-2.5U/50µl
Water, nuclease-free	to 50 µl	–

Recommendation amounts of template DNA in a 50 µl reaction mix:

Human genomic DNA	0.1µg-1µg
Plasmid DNA	0.5ng-5ng
Phage DNA	0.1ng-10ng
E.coli genomic DNA	10ng-100ng

2. Mix contents in the tube. Cap tubes and centrifuge briefly to collect the contents to the bottom.

When using a thermal cycler that does not contain a heated lid, overlay the reaction mixture with 25 µl mineral oil.

3. Perform 25-35 cycles of PCR amplification as follows:

Initial Denaturation	94°C	3 minutes
25-35 Cycles	94°C	30 seconds
	55-68°C 72°C	30 seconds 1-5 minutes
Final Extension	72°C	2 minutes

4. Incubate for an additional 2 min at 72°C and maintain the reaction at 4°C. The samples can be stored at -20°C until use.

5. Analyze the amplification products by agarose gel electrophoresis and visualize by nucleic acid dye staining. Use appropriate molecular weight standards.

Notes on cycling conditions

- *FS*TM Taq DNA Polymerase is for high specificity PCR applications.
- The half-life of enzyme is >40 minutes at 95°C.
- The error rate of *FS*TM Taq DNA Polymerase in PCR is 2.2×10^{-5} errors per nt per cycle.
- *FS*TM Taq DNA Polymerase accepts modified nucleotides (e.g. biotin-, digoxigenin-, fluorescent-labeled nucleotides) as substrates for the DNA synthesis.
- The number of PCR cycles depends on the amount of template DNA in the reaction mix and on the expected yield of the PCR product. 25-35 cycles are usually sufficient for the majority PCR reaction. Low amounts of starting template may require 40 cycles.

Guidelines for preventing contamination of PCR

During PCR more than 10 million copies of template DNA are generated. Therefore, care must be taken to avoid contamination with other templates and amplicons that may be present in the laboratory environment. General recommendations to lower the risk of contamination are as follows:

- Prepare your DNA sample, set up the PCR mixture, perform thermal cycling and analyze PCR products in separate areas.
- Set up PCR mixtures in a laminar flow cabinet equipped with an UV lamp.
- Wear fresh gloves for DNA purification and reaction set-up.
- Use reagent containers dedicated for PCR. Use positive displacement pipettes, or use pipette tips with aerosol filters to prepare DNA samples and perform PCR set-up.
- Always perform “no template control” (NTC) reactions to check for contamination.

Quality Control

The absence of endodeoxyribonucleases, exodeoxyribonucleases and ribonucleases is confirmed by appropriate quality tests. Functionally tested in amplification of a single-copy gene from human genomic DNA.

Endodeoxyribonuclease Assay

No detectable conversion of covalently closed circular DNA to a nicked DNA was observed after incubation of 10U *FS*TM Taq DNA Polymerase with 1µg pBR322 DNA for 4 hours at 37°C and 70°C.

Exodeoxyribonuclease Assay

No detectable degradation of lambda DNA-HindIII fragments was observed after incubation of 10U *FS*TM Taq DNA Polymerase with

1 μ g digested DNA for 4 hours at 37°C and 70°C.

Ribonuclease Assay

0% of the total radioactivity was released into trichloroacetic acid-soluble fraction after incubation of 10U *FS*TM Taq DNA Polymerase with 1 μ g E.coli [3H]-RNA (40000cpm/ μ g) for 4 hours at 37°C and 70°C.